

NGS technology & applications – the past, present & future

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What is sequencing, and why bothering?

Sequencing = A technique for determining the exact sequence of nucleotides, or bases, in a DNA or RNA molecule.



"... [A] knowledge of sequences could contribute much to our understanding of living matter."

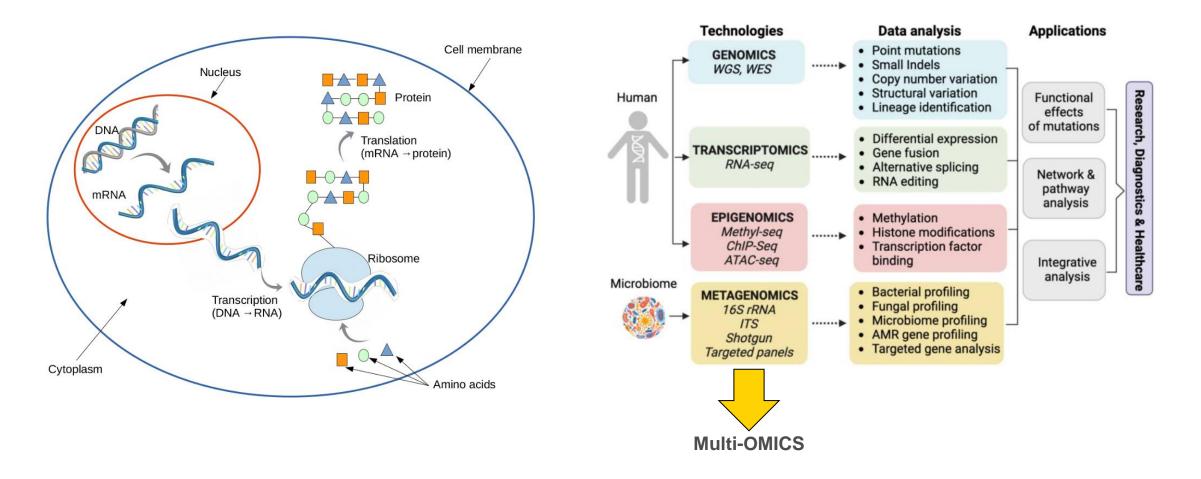
[Frederick Sanger]

- 2 Noble prices
 - determining the amino acid sequence of insulin and other proteins
 - The Sanger (chain-termination) method for DNA sequencing
 - One of the first to sequence RNA (5S rRNA) in 1967 (Robert W. Holley and team sequenced first RNA (S. cerevisae Ala-tRNA in 1965)



What is sequencing, and why bothering?

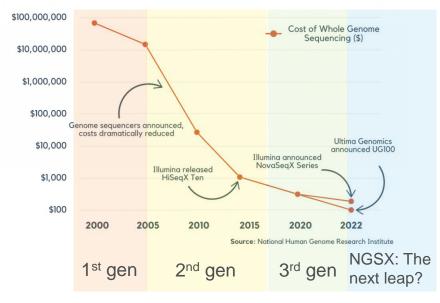
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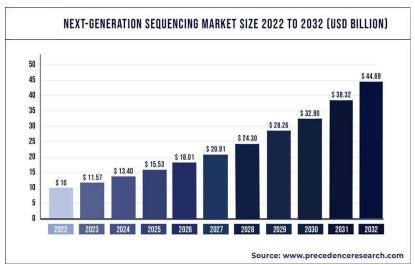


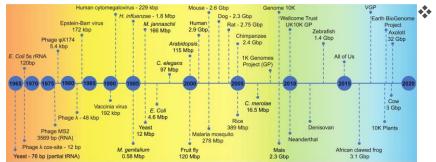
Sequencing on its way to be democratized

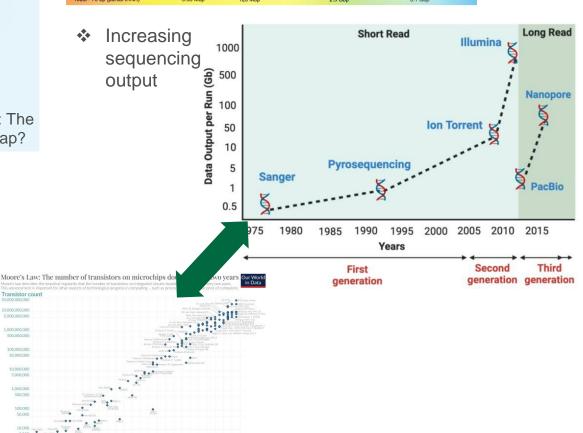
 Decreasing genome sequencing costs



 Increasing market size



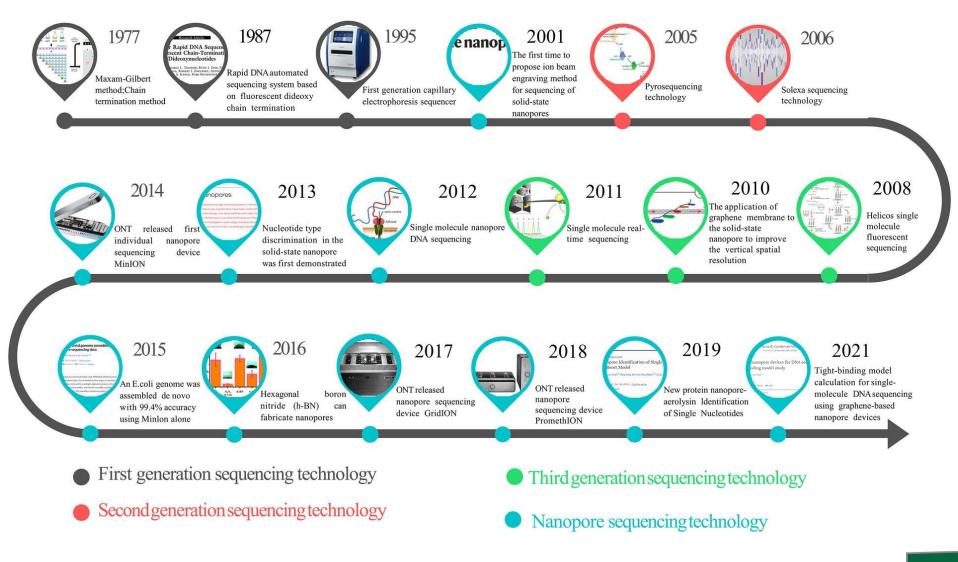




Milestones



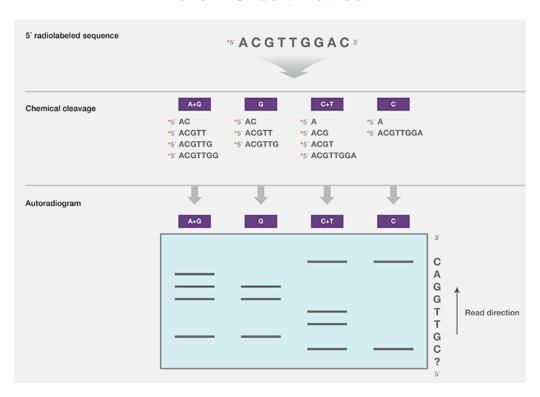
Evolution of sequencing technologies



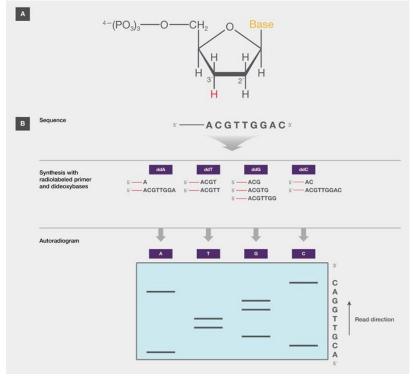


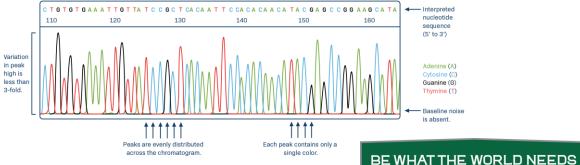
Classical sequencing – the gold standard

Maxam-Gilbert method



Sanger method







The beginnings of NGS technology

Pyrosequencing

Principle in 1993 (Pal Nyren et al. U Stockholm), first commercial sequencer by 454 Life Sciences in 2004

Principle

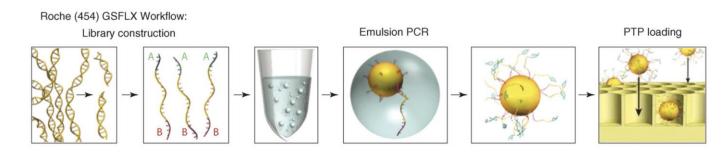
- Sequencing by synthesis
- Utilizes pyrophosphate to detect nt incorporation
- Parallel sequencing
- First NGS technology

Advantages

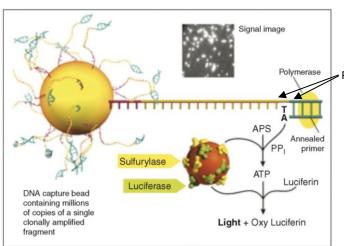
- High throughput compared to Sanger (600Mbp/10hrs vs 2Mbp/day)
- Real-time monitoring
- No electrophoresis
- Suitable for SNP and indel detection

Disadvantages

- Short read length (100-300bp vs 500-1000bp)
- Poor sequencing capacity for homopolymer, repetitive and GC-rich regions







Pyrosequencing reaction

Phosphodiester bonds

Nucleotide sequence
G C - A GG CC T

G C T A G C T

Nucleotide added

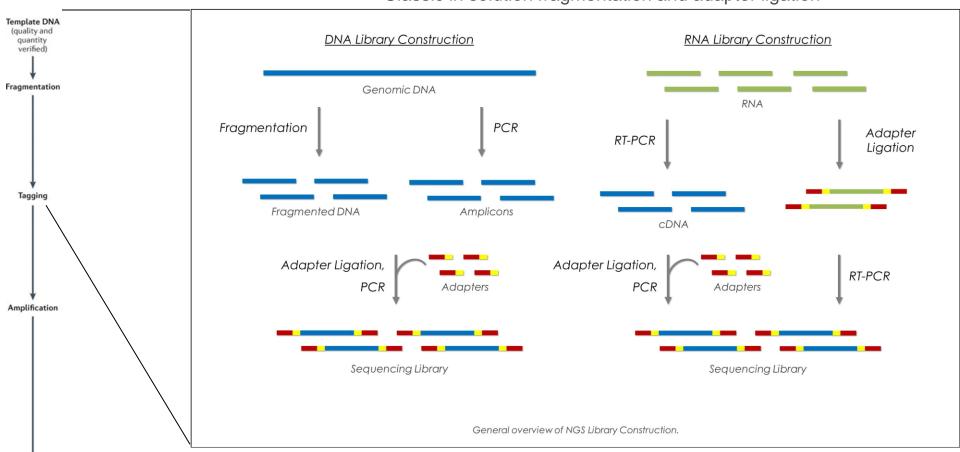
TRENDS in Genetics



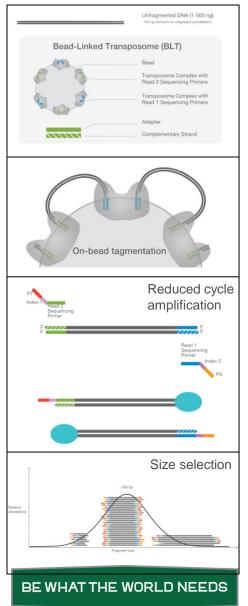
The 2nd generation: NGS/shotgun/massive

parallel sequencing

Classic in solution fragmentation and adapter ligation



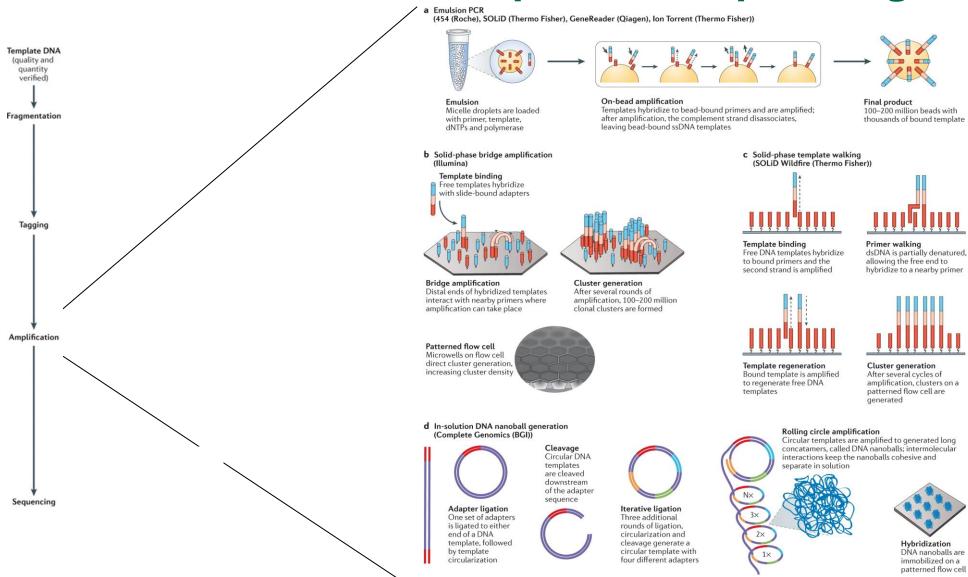
On bead Tagmentation



Sequencing



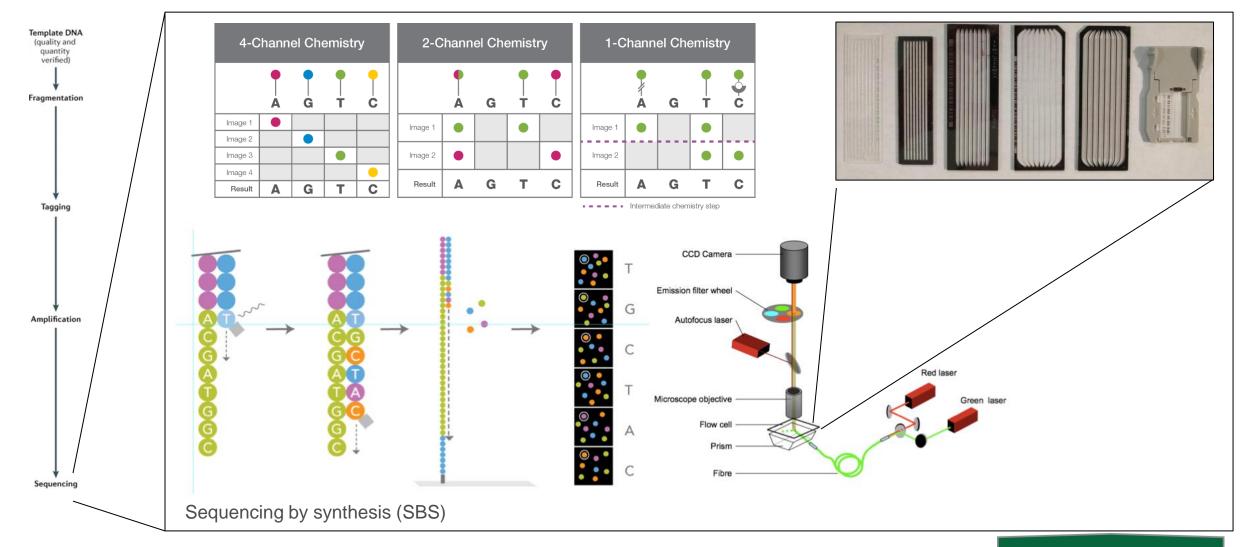
The 2nd generation: NGS/shotgun/massive parallel sequencing



BE WHAT THE WORLD NEEDS

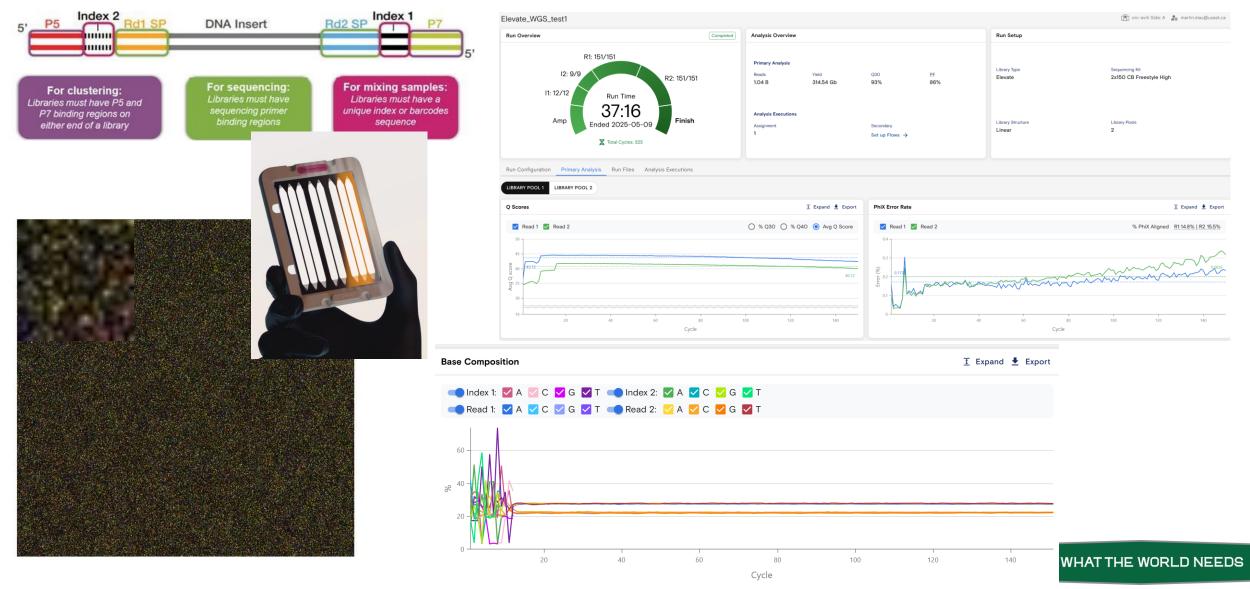


The 2nd generation: NGS/shotgun/massive parallel sequencing





The 2nd generation: NGS/shotgun/massive parallel sequencing





3rd Generation Sequencing - ONT

Technology

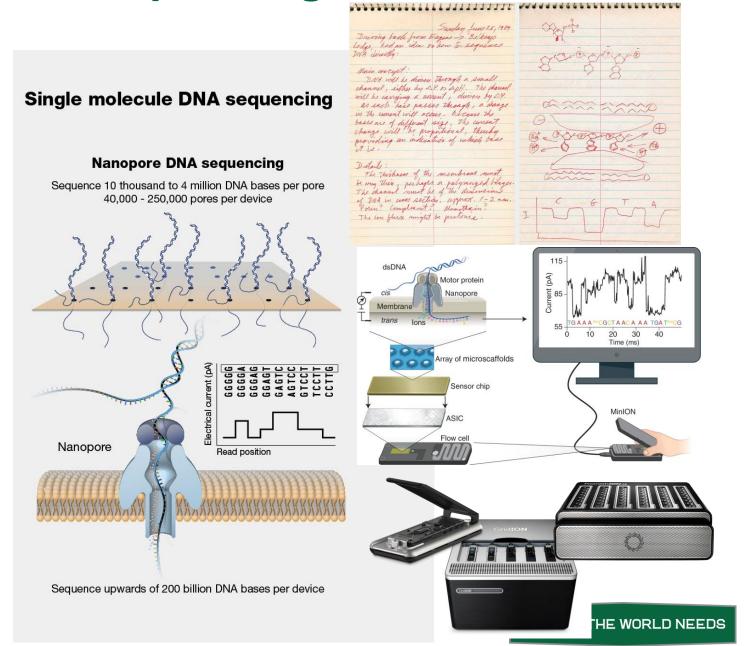
- ❖ Beginnings in 1989, seminal study in 2018
- Electrical current, drives an unwound oligomers (RNA/DNA) through an immobilized protein nanopore channel, disrupting the current as they passed through, in a manner characteristic of their base composition.
- Detection of electrical signals not fluorescence

Advantages

- Read limitations only by length of HMW DNA (Read length N50: 100-200kbp → 4Mbp)
- De novo genome assemblies
- Genomic + Epigenomic information
- Field sequencing

Disadvantages

- Lower accuracy (Phred Q score ~20)
- Lower output in gigabases (10s Gb/FC)

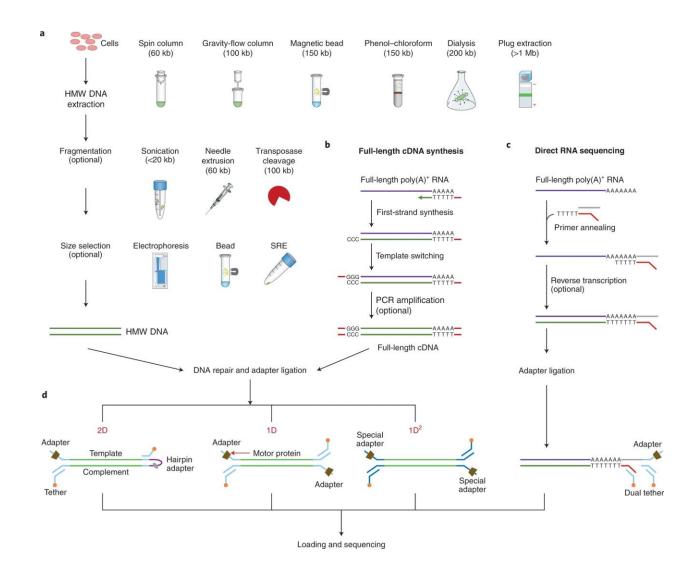




3rd Generation Sequencing - ONT

Sample preparation

- HMW DNA extraction is sensitive process
- QC requires specific equipment (Femtopulse)
- 2D (library for template + complement strand sequencing)
- 1D (each strand ligated and sequenced separately)
- 1D2 (sequential sequencing of both strands)

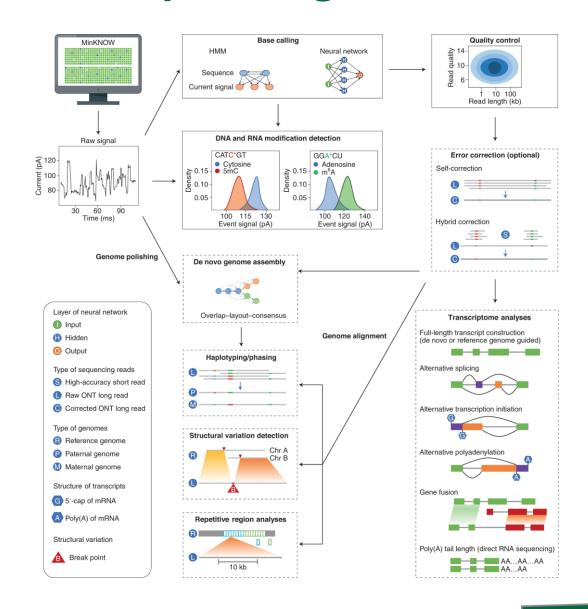




3rd Generation Sequencing - ONT

ONT data ideal for

- Multiploid organisms
- Complex genomes (e.g. recurrent hybrids, young and fast evolving genomes, genomes with high plasticity)
- Organisms without reference genomes or close relatives with reference genomes (de novo assembly)
- Detection of 5-Methylcytosin (5mC) modifications





OMICS 3rd Generation Sequencing – SMRT-Seq

Technology

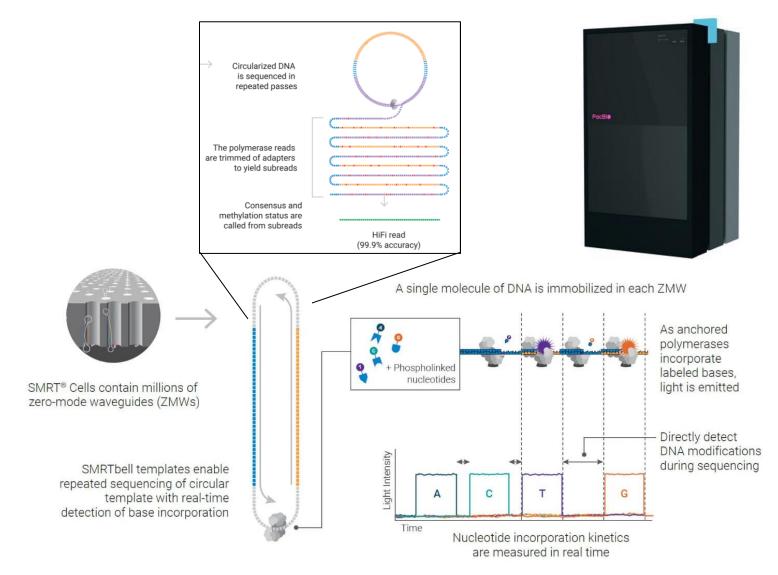
- Single molecule sequencing in ZMWs
- SBS chemistry and photometrics

Advantages

- Merged reads from multiple passes enables higher accuracy compared to ONT (Q30+)
- Read length N50: 10-60kb
- Genomic + Epigenomic information (5mC in CHG context)

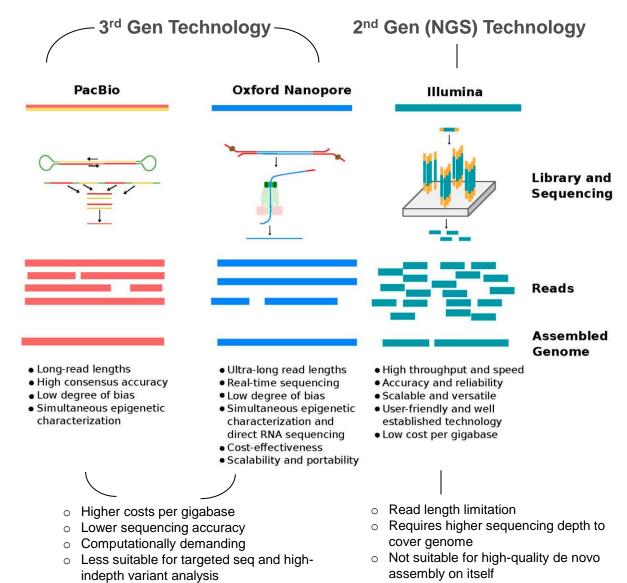
Disadvantages

Higher costs compared to ONT sequencing (\$43-86/Gb vs. \$21-42/Gb)





Sequencing technologies



BE WHAT THE WORLD NEEDS

o Requires larger input and high-quality

samples

Advantages

Disadvantages



Changes to sample prep: scRNA-Seq

Technology

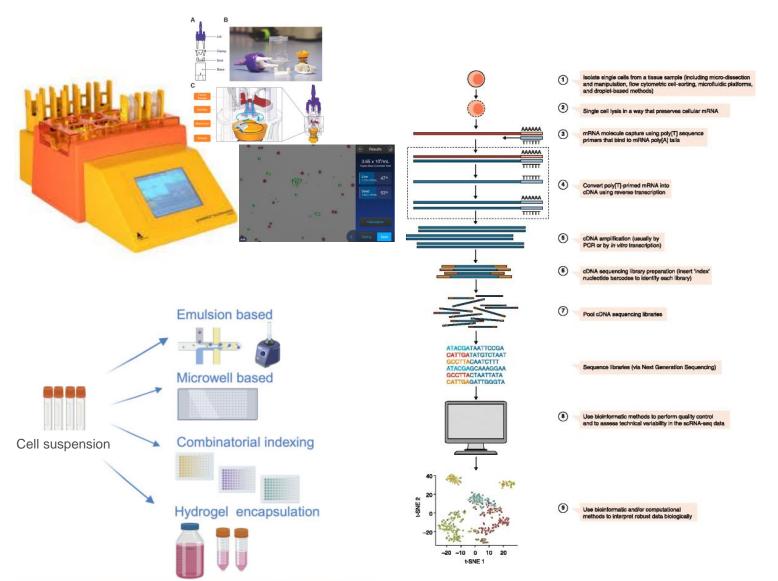
- various
- Utilizes any standard NGS technologies
- 10-50k reads per cell & 1k-mio cells

Advantages

- comparison of the transcriptomes of individual cells
- Addresses tissue heterogeneity (e.g. embryonic or immune cells)
- Identification of rare cell populations (e.g. normal-tumor interfaces)
- Characterization of unique cells (e.g. Tlymphocytes, neurons)
- Cell fate diversification

Disadvantages

- High costs
- Computation intense





Changes to sample prep: Spatial sequencing

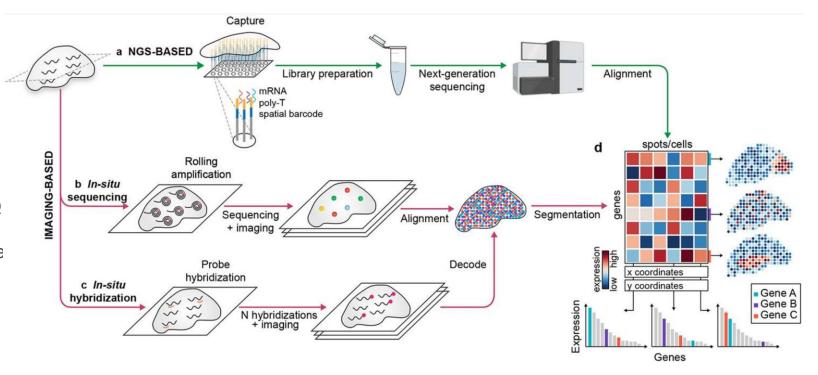
Technology

- Combines FISH and standard NGS for spatial genomics (clusters - not single cell)
- Commercial ISS technologies:
 - 10X Genomics Xenium & Visium
 - Curio Bio SEEKER

Approaches

- NGS-based (Slide-Seq, DBiT-Seq)
- ISS (HybISS: Hybridization-based in situ sequencing (Nilsson lab, U Stockholm), FISSEQ fluorescent in situ RNA sequencing (Church lab, Wyss Institute), INSTA-Seq: in situ transcriptome accessibility sequencing (Lee lab, CSH)
- ISH (MERFISH, SeqFISH)

- Cell type identification
- Tumor-immune composition
- Cellular-molecular organization
- Tracking cell fate/development





OMICS Changes to sample prep: In situ sequencing (ISS)

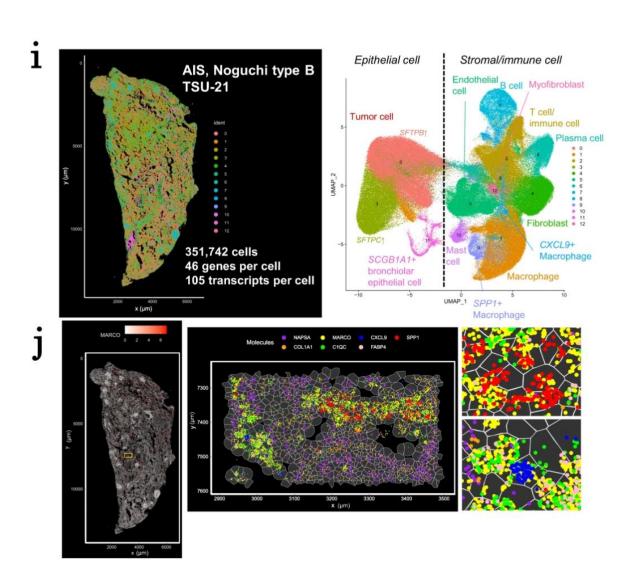
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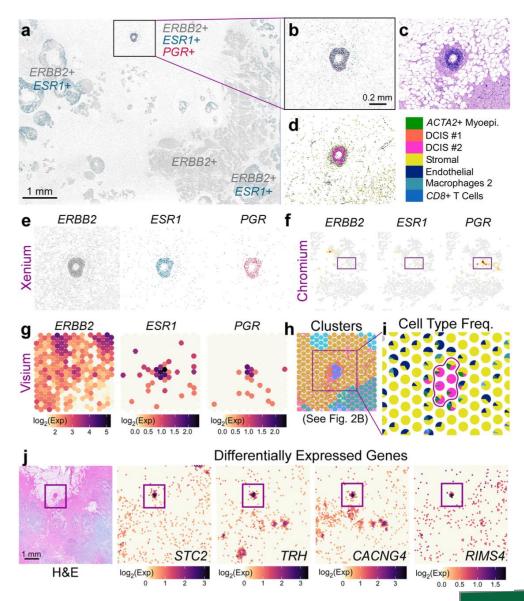
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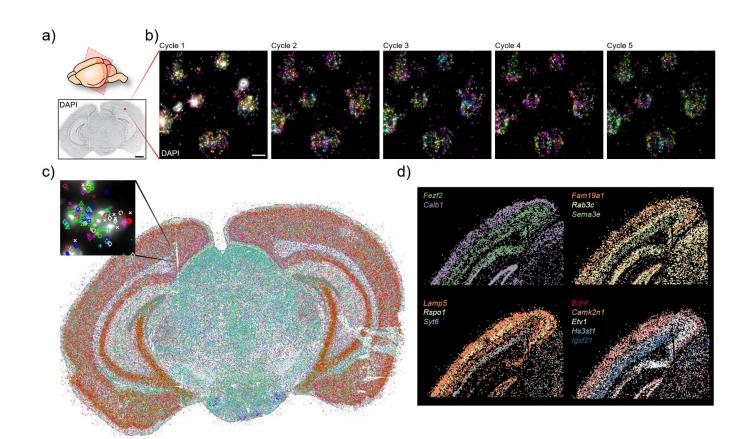
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- Cell type identification
- Tumor-immune composition
- Cellular-molecular organization
- Spatial mapping of transcript activity
- Tracking cell fate/development



Mouse coronal brain section



OMICS Changes to sample prep: In situ sequencing (ISS)

Technology

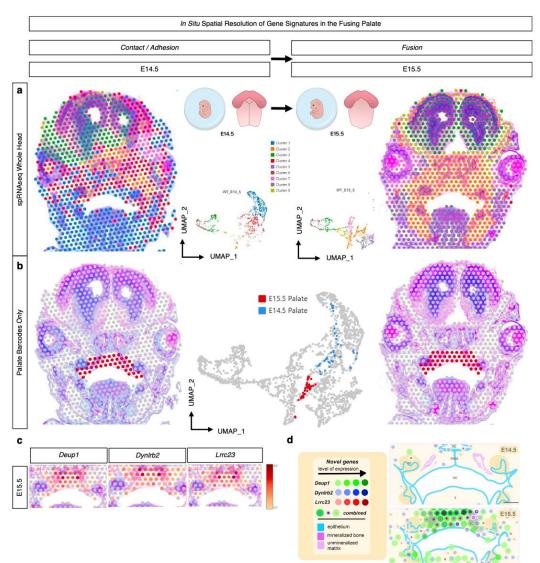
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Applications

- Cell type identification
- Tumor-immune composition
- Cellular-molecular organization
- Spatial mapping of transcript activity
- Tracking cell fate/development, e.g. to understand developmental disorders



Whole human embryo heads



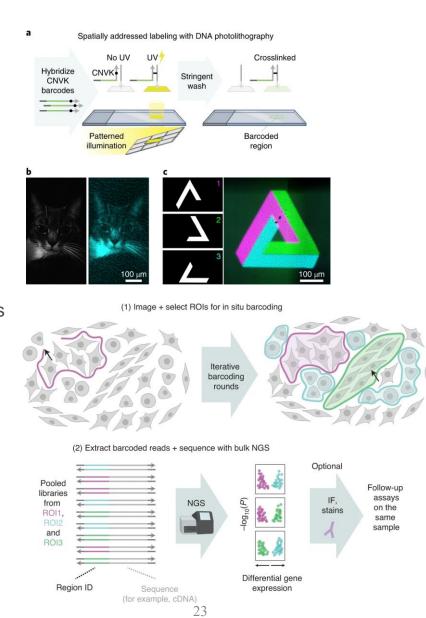
Changes to sample prep: Light-Seq

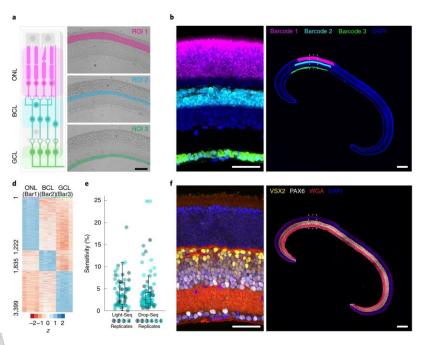
Technology

- photocrosslinkable nucleosides analog to 'geotag' the transcriptome of target cells in situ by light-controlled attachment of DNA barcodes
- high resolution imaging with standard NGS

Advantages

- Profiling of multiple different cell populations in fixed biological samples
- Spatial resolution of gene expression
- microscopically analyzed cells kept intact during RNA expression profiling which allows sequential measurements







Changes to the hardware: The spinning disc

Off-the-shelf semiconductor hardware

spinning silicon wafer substitute flow cells

Traditional chemistry

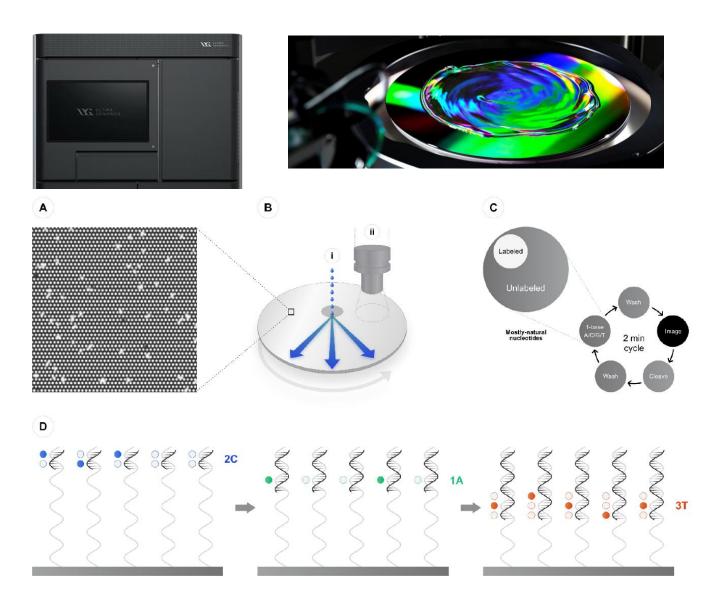
natural sequencing-by-synthesis (mnSBS)

Open-fluidics-system and fixed optics

 Sequencing reagents are added via spincoating, and for readout of DNA sequence, the silicon wafer rotates while fixed dual highspeed cameras capture DNA clusters immobilized on the wafer.

Advantages:

- ✓ Ultra-high-throughput and ultra-low costs (\$100 Genome)
- ✓ Short run-times, e.g. six human genomes in 20hrs
- ✓ Longer average read lengths (300-400+ cycles)





Changes to the chemistry: ABC-sequencing



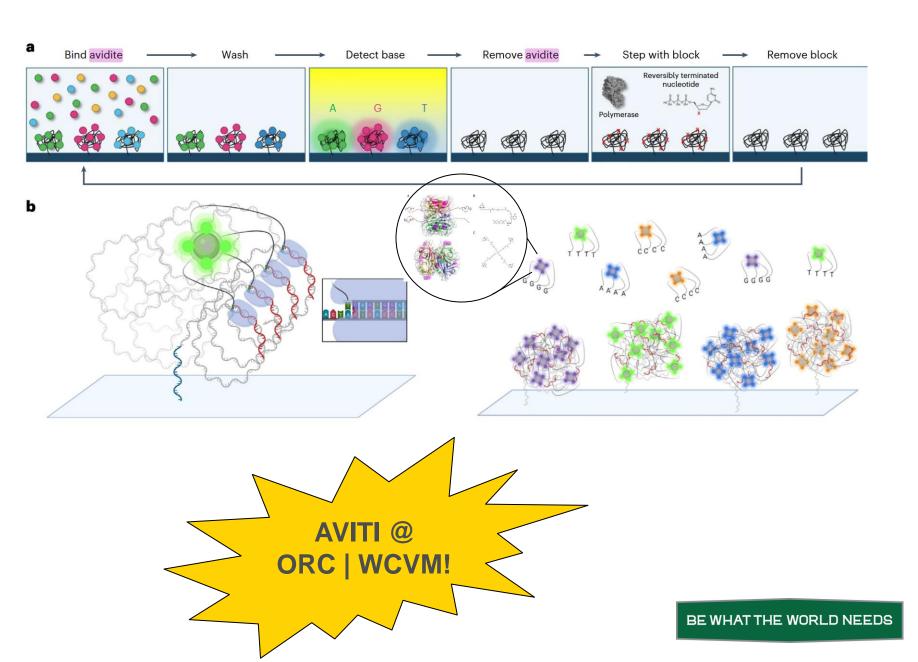


Changes to Illumina's SBS chemistry

- Circularization of the DNA library molecule on a capture primer
- RC-Amplification copies ccDNA into a continues strand bound into a polony
- Sequencing using Avidites

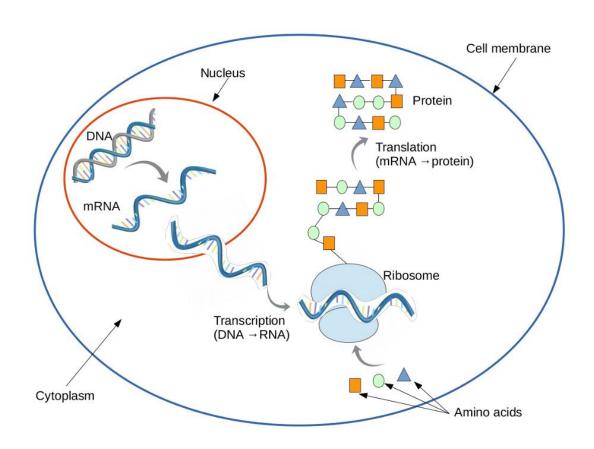
Advantages

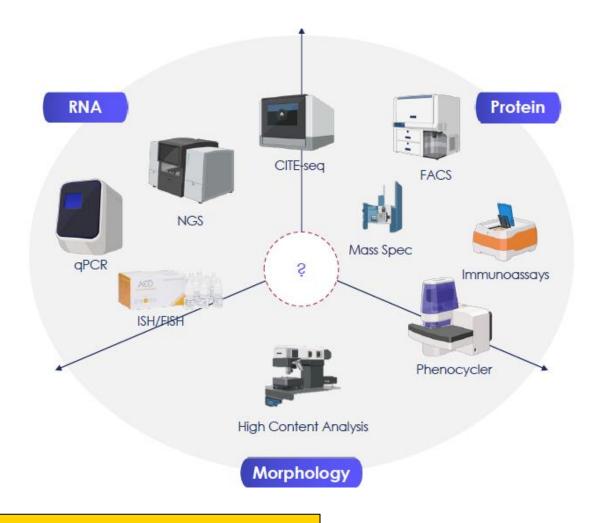
- ✓ No PCR artifacts
- ✓ Reduced error rate
- ✓ Reduced costs
- √ No index hopping





The "Future" of sequencing: NGSX





Integration of true multiomics capabilities on a single instrument without the need of auxiliary equipment



Direct in sample sequencing (DISS) for cells

Technology:

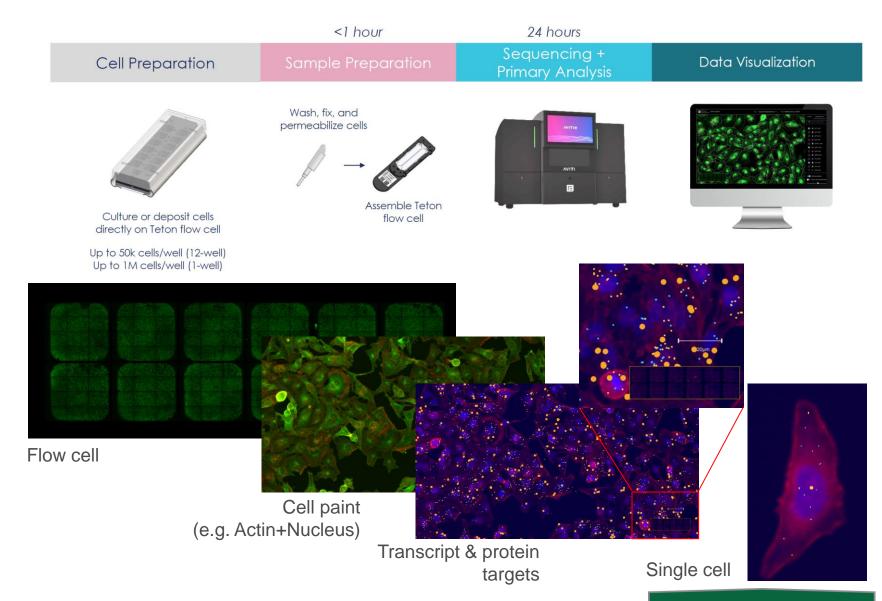
 Teton CytoProfiling + ABC sequencing from Element Biosciences

True multiomics on a single instrument

- Cell morphology
- RNA detection (~350-plex)
- Protein detection (~50-plex)

Use cases:

- Cellular profiling
- drug response studies
- o in situ single cell RNA-Seq
- cell synchronization





DISS for FFPE & fresh tissue

Technology:

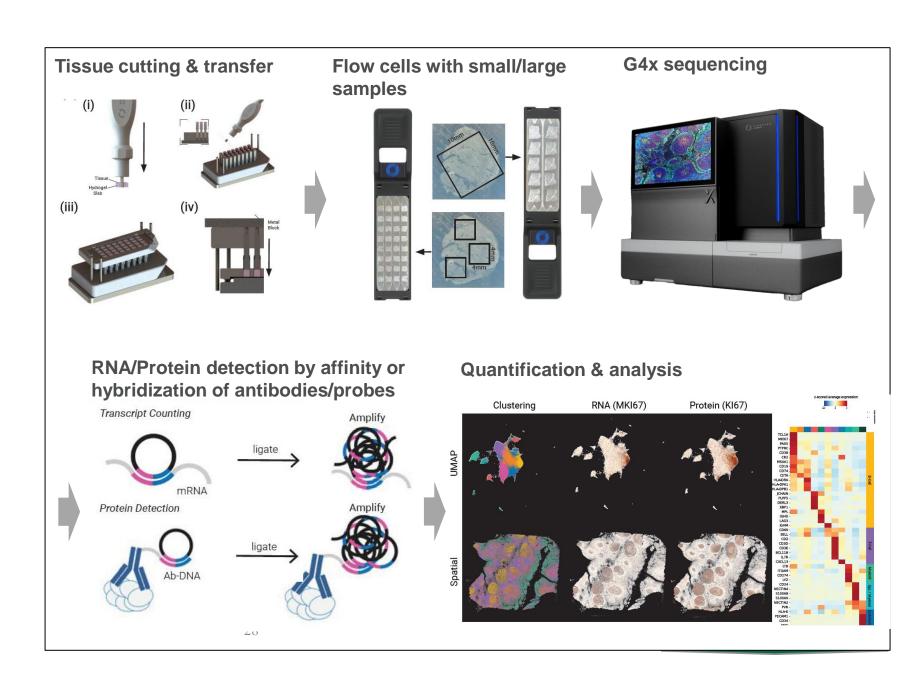
Tissue transfer + rapid 4-color
 SBS sequencing by Singular
 Genomics

True multiomics on a single instrument

- fH&E (Hematoxylin and eosin)
- RNA detection (~153-plex)
- Protein detection (~10-plex)

Use cases:

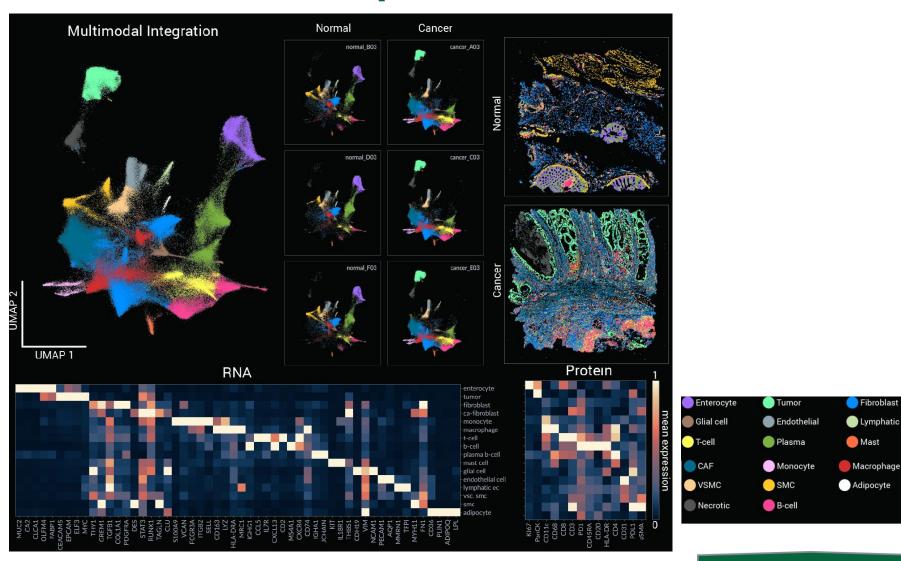
- late-stage translational research (T2-T4); e.g. tumor-immune interactions
- Diagnostics; e.g. therapeutic biomarkers





DISS use case #1: Visualize crosstalk of multiple cellular processes

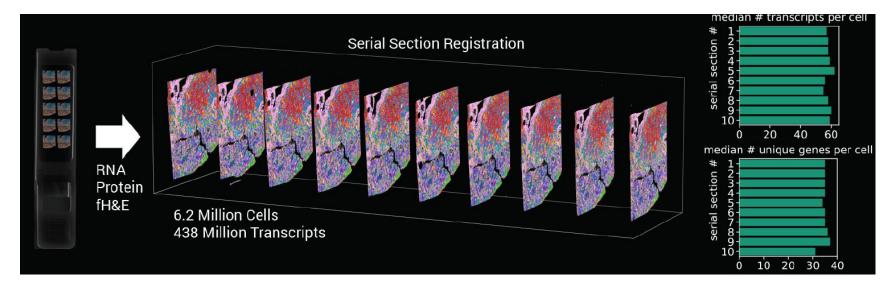
- Tumor-Immune interactions (e.g. Colon normal vs cancer)
- Visualize & sequence tissue with (Sub-)Cellular resolution
- Targeting 100 millions of transcripts in millions of cells
- Integration of complex single cell RNA & protein expression profiles

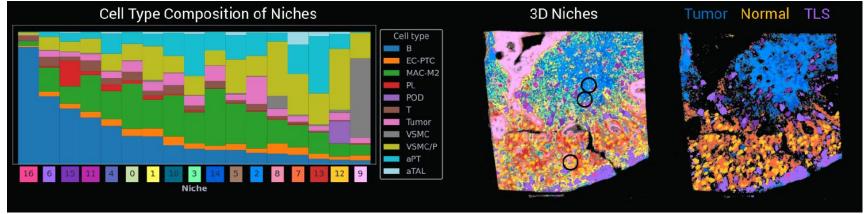




DISS use case #2: Enhance therapeutic success

- Reconstruction of Normal-Tumor Interfaces
- 3D spatial neighborhood analysis to identify niches of healthy vs cancerogenous tissue
- Monitoring effects of tumormodulating drugs e.g. to enhance effectiveness of chemotherapies



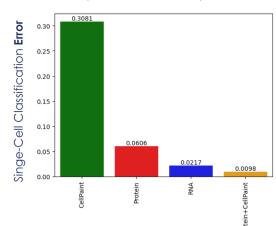


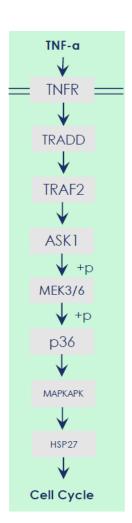


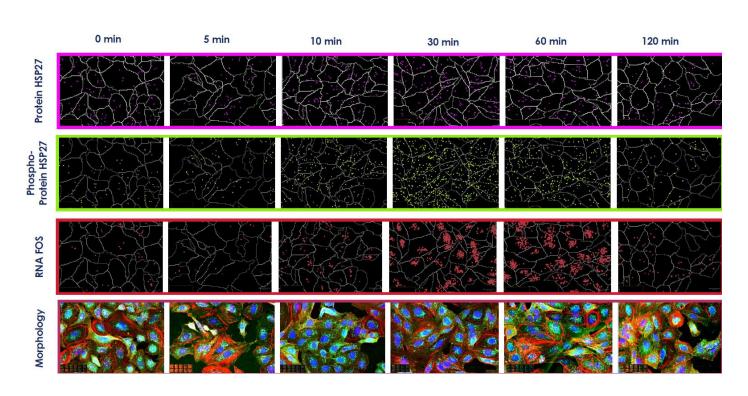
DISS use case #3: Personalized drug therapy

- Drug-induced changes to RNA, protein, phosphoprotein, and morphology
- Multiomic readouts identifies RNAs and Proteins driving the changes of morphological states & resolves complex mechanisms of drug response
- Multiomic readouts improve classification of cells based on their drug response

TNF-alpha treatment shows a strong protein and RNA response









Summary & Trends

- ✓ Sequencing is a fast-evolving field of new emerging technologies
- ✓ New NGS methods are faster and more precise
- ✓ Necessitate smaller amounts of input DNA and reagents
- ✓ NGS technologies created new fields of research (e.g. pangenome analysis, multiomics, cell atlases)
- True multi-omics revolutionizes basic & translational research



How to handle the enormous amount of data ("big data")?



Development of new bioinformatic tools, statistics, employ data sharing, unified file formats and AI to help integrate data from different "omes"



The Omics Resource Centre (ORC)







